BUDDLEDIN A, B AND C, PISCICIDAL SESQUITERPENES FROM BUDDLEJA DAVIDII FRANCH.

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Some plants of *Buddleja* species have been known to be poisonous to fishes. We have isolated from the root bark of *Buddleja davidii* Franch. (Buddlejaceae), three new piscicidal sesquiterpenes, buddledin A (1), B (2) and C (3).

Chromatography over silica gel followed by preparative TLC guided by the killie-fish test¹, gave buddledin A, B and C in ca. 0.5, 0.003 and 0.03% yields (from the fresh root bark), whose spectral data are listed in Table 1.

Buddledin A, $C_{17}H_{24}O_3$ (M⁺ 276), m.p. 94-95°, $[\alpha]_n$ -245° (c, 0.74, CHCl₃), shows in the spectra the presence of an α , β -unsaturated ketone substituted by CH₂ at α - and by CH₂ at β -carbon (UV 238 nm, IR, 1682, 1642 cm⁻¹, PMR, β-H at δ 6.45 (m) coupled with α -CH₃ at δ 1.64 (d, J=2 Hz). An acetyl group (IR, 1742, 1236 cm⁻¹, PMR, δ 2.14) on a secondary carbon whose proton (H_A, δ 5.67, d, J=11 Hz, shifted to δ 4.69 on deacetylation of 1), is coupled with another methine proton (H_p, t, J=11 Hz) which is resolved by addition of Eu(fod), is also exhibited. A two-proton singlet at δ 4.93 of exomethylene group (IR, 890 cm⁻¹), replaced by a secondary methyl signal at δ 0.83 (d, J=7 Hz) upon hydrogenation of 1 to tetrahydrobuddledin A (4), C17H2803 (M⁺ 280), m.p. 88-89°, is shown by addition of the shift reagent to be coupled with allylic H_c (br.q) which is coupled with $H_{B}^{}$. Two tertiary methyl groups (PMR, δ 1.11 and 1.10, CMR², two of three signals at δ 13.7, 20.6 and 21.6) on a quaternary carbon (CMR, δ 34.0), and two additional methylene carbons (CMR, two of three peaks at δ 30.0, 40.0 and 41.3) are also exhibited. The gem-dimethyl-carrying carbon and one of the methylene groups are presumed to be located adjacent to H_p - and H_c -carrying carbon, respectively, on the basis of the splitting patterns of these protons $(H_{p}, t, and$ H_c , br.q). The AcO-carrying carbon is shown to be α to the ketone group by the LiAlH₄ reduction of 1 to epimeric alcohols, 5, C15H24O2 (M⁺ 236), m.p. 123-125°, and 6, m.p. 70-72°, which show coupling of newly formed H_D with H_A [5, H_A δ 3.75 (dd, J=2, 10 Hz), H_D δ 4.17 (d, J=2 Hz); 6, H_A δ 3.74 (t, J=9 Hz), H_n δ 3.50 (d, J=9 Hz)]. The couplings in these PMR data were confirmed by PMDR. These spectral data indicate partial structure la, and coupled with the biogenetic considerations, lead to structure 1 for buddledin A.

The structure and relative stereochemistry including the geometry of the endocyclic double bond as depicted in <u>1</u> were unequivocally established by X-ray diffraction analysis of the bromohydrin <u>7</u>, m.p. 179-180°, which was derived from <u>1</u> by the treatment with N-bromoacetamide in aqueous acetone. Clear crystals of <u>7</u> belong to the orthorhombic space group $P2_12_12_1$ with unit cell axes a=19.747, b=8.693, c=10.194 Å, and four molecules of $C_{17}H_{25}O_4Br$ per unit cell. Of 1879 reflections, collected with a Syntex P; automated diffractometer, 1133 were above the background (2 σ). The structure was solved by the heavy atom method and refined by the full-matrix least-squares method to a conventional R-value of 0.078 without hydrogen atoms.

The absolute configuration as given in <u>1</u> was determined by the application of dibenzoate chirality rule³ and benzoate rule⁴ as follows. CD spectra of dibenzoate <u>8</u>, $C_{29}H_{92}O_4$ (M⁺ 444), m.p. 129-130°, PMR, δ 5.36 (dd, J=3, 10 Hz, C₂-H), 5.67 (d, J=3 Hz, C₃-H), and <u>9</u>, $C_{29}H_{32}O_4$ (M⁺ 444), m.p. 122-123°, PMR, δ 5.64 (t, J=10 Hz, C₂-H), 5.31 (d, J=10 Hz, C₃-H), obtained from two epimeric diols <u>5</u> and <u>6</u>, show positive ($\Delta \varepsilon_{236}$ +20.5, $\Delta \varepsilon_{220}$ -3.5) and negative ($\Delta \varepsilon_{237}$ -15.25, $\Delta \varepsilon_{223}$ +19.0) chiralities, which indicate β orientation of C₂-OH. Application of the benzoate rule for monoalcohol <u>13</u>, C₁₅H₂₈O (M⁺ 224), [M]_D +13.2°, derived from <u>4</u> by four-steps sequence (<u>4</u> \rightarrow <u>10</u> \rightarrow <u>11</u> \rightarrow <u>12</u> \rightarrow <u>13</u>), and its benzoate <u>14</u>, C₂₂H₃₂O₂ (M⁺ 328), [M]_D +207.8°, agrees with the result from the dibenzoate chirality rule.

Buddledin B, $C_{15}H_{22}O_2$ (M⁺ 234), m.p. 139-141°, $[\alpha]_D$ -314° (c, 0.54, CHCl₃), was proved to be deacetylbuddledin A (<u>2</u>) obtainable by deacetylation of <u>1</u>. Ethyl ether <u>15</u>, $C_{17}H_{20}O_3$ (M⁺ 280), m.p. 95-96°, along with <u>2</u>, was produced upon the ethanolysis. Acetylation of <u>2</u> regenerated buddledin A.

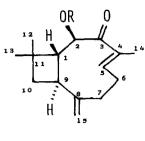
The spectroscopic data of buddledin C, $C_{15}H_{22}O(M^+ 218)$, m.p. $129-130^\circ$, $[\alpha]_D^- -316^\circ$ (c, 0.54, CHCl₃) suggest it to be deacetoxybuddledin A (<u>3</u>) and this has been confirmed as follows. With Zn-AcOH, buddledin A was converted to deacetoxydihydrobuddledin A (<u>16</u>), $C_{15}H_{24}O(M^+ 220)$, $[\alpha]_D^+ +42.6^\circ$ (c, 0.27, dioxane), and dihydrobuddledin A (<u>17</u>), $C_{12}H_{26}O_3(M^+ 278)$. The former was

Compound	V ^{KBr} , cm ⁻¹	λ ^{MeOH} , nm PMR (90 MHz, CDCl ₃ , δ in ppm, J in Hz)					
		max - (ε)	н-2	H-5	H-12, 13	н-14	H -1 5
<u>1</u>	1742, 1682, 1642	238	5.67	6.45	1.11 (s)	1.64	4.93 (s)
	1236, 890	(8,500)	(d, 11)	(m)	1.10 (s)	(d, 2)	
<u>2</u>	3440, 1655, 1635	237	4.69	6.42	1.29 (s)	1.69	4.90 (s)
	1615, 903	(9,400)	(q, 5, 11) ^a	(m)	1.09 (s)	(d, 2)	
3	1662, 1637, 887	236	3.0-1.5 ^b	6.33	1.00 (s)	1,63	4.92 (s)
		(11,000)		(m)		(d, 2)	4.95 (s)
<u>18</u>	1667, 880	235	2.7-1.5 ^b	5.53	1.02 (s)	1.77	4.88 (br.s)
		(4,700)		(m)	1.01 (s)	(q, 1~2)	4.82 (br.s)
<u>19</u>	1738, 1707, 1668	237	5.27	5.60	1.13 (s)	2.01	4.85 (s)
	1633, 1236, 895	(3,500)	(d, 11)	(m)	1.09 (s)	(d, 1)	

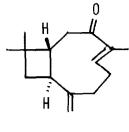
Table 1. Spectral Data of 1, 2, 3 and Their Derivatives

(a) Transformed to a doublet (J=11 Hz) on addition of D₂O.

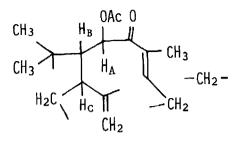
(b) Overlapped by other protons.



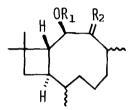
 $\frac{1}{2} R = H$



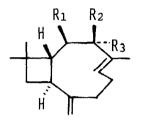
<u>3</u>

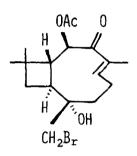


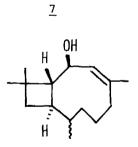
<u>la</u>



 $\begin{array}{cccc} \underline{4} & R_1 = Ac, & R_2 = 0 \\ \underline{10} & R_1 = Ac, & R_2 = \checkmark {}_{H}^{OH} \\ \underline{11} & R_1 = Ac, & R_2 = \checkmark {}_{H}^{OMs} \\ \underline{13} & R_1 = H, & R_2 = H_2 \\ \underline{14} & R_1 = Bz, & R_2 = H_2 \end{array}$



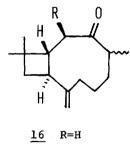




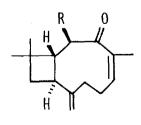
12

H H H H H OH OEt

15



<u>17</u> R=OAc



<u>18</u> R=H

<u>19</u> R=OAc

identified with the product (dihydrobuddledin C) obtained by the same treatment of buddledin C. On Zn-AcOH reduction in diluted solution in a shorter time, <u>3</u> afforded, in addition to <u>16</u>, an isomer <u>18</u>, $C_{15}H_{22}O$ (M⁺ 218), $[\alpha]_{D}$ -199° (c, 0.7, CHCl₃), which was converted to <u>16</u> on prolonged reaction. This isomer was also obtained together with <u>16</u> and <u>17</u> by an analogous reaction of <u>1</u>. Buddledin A was slowly isomerized in MeOH or Me₂CO solution at room temperature to give a *cis* isomer <u>19</u>, $C_{17}H_{24}O_3$ (M⁺ 276). This isomerization occurred quickly by the UV irradiation of <u>1</u> in *n*-hexane, as found with caryophyllene⁵. The weak UV absorption of α , β -unsaturated ketone in <u>19</u> and the upfield shift of C₅-olefinic proton in PMR spectrum would be due to torsion of the conjugated system in the *cis* isomer. The isomerization of <u>3</u> + <u>18</u> has been indicated to be from *trans* to *cis* by the analogous change in the spectra (Table 1).

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